



Linking Seasonal Foliar Chemistry to VSWIR-TIR Spectroscopy Across California Ecosystems

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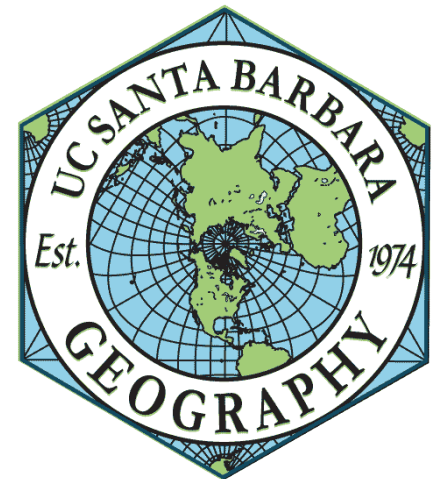
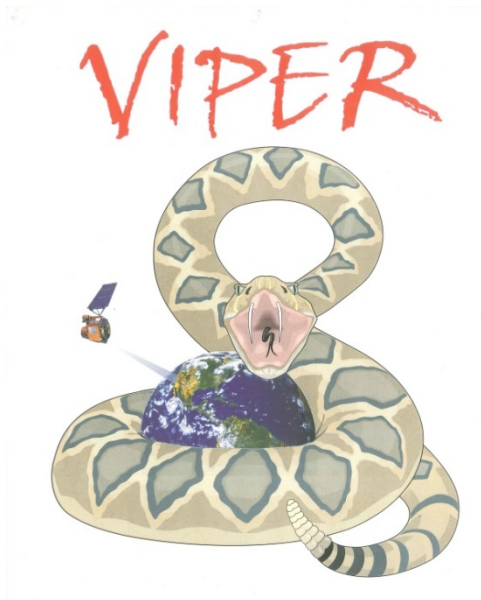
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Foliar Chemistry

Foliar chemistry provides an insight into the quality of vegetation and nutrient cycles

- Chlorophyll → Levels of productivity
- Nitrogen → Indicator of photosynthetic and growth rates
- Lignin → Rate of litter decomposition

Conventional Methods are time consuming and expensive

Spectroscopy can address these issues through faster analytical speed and minimal sample preparation (Lawler et al., 2006)

VSWIR Spectroscopy

- Most spectroscopy studies use the Visible Near Infrared/Short Wave Infrared (VSWIR) spectrum
- Plant Biochemical:
 - Nitrogen (Bolster et al. 1996; Doughty et al 2009; Ferwerda et al., 2005; Martin et al. 2008)
 - Lignin and Cellulose (Kokaly and Clark, 1992; Asner et al., 2011; Kokaly et al., 2009)
 - Chlorophyll (Ustin et al., 2009; Doughty et al 2009; Martin et al.,2007; Asner et al., 2011)
- Plant Biophysical: Specific Leaf Area, Water Content
 - (Ceccato et al., 2001; Doughty et al., 2009; Martin et al., 2007; Asner et al., 2011)
- However, dominated by water and pigment absorption (Ribeiro da Luz and Crowley, 2010)

Spectroscopy using TIR

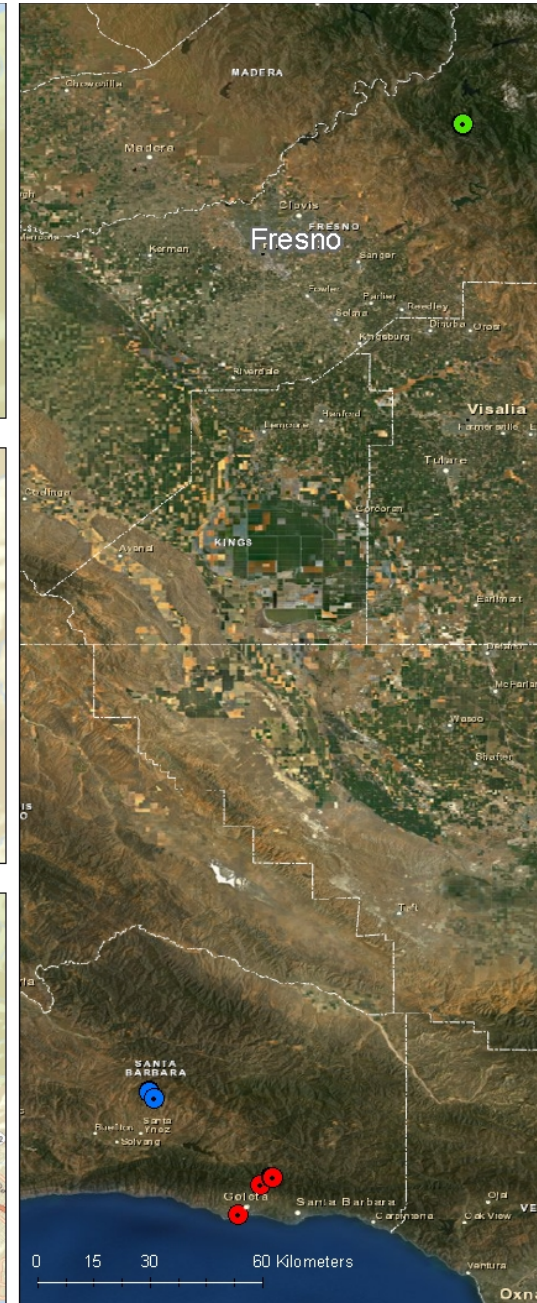
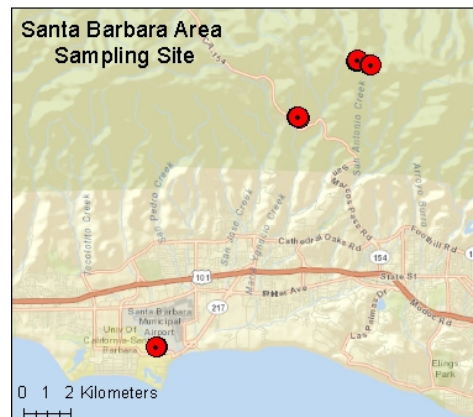
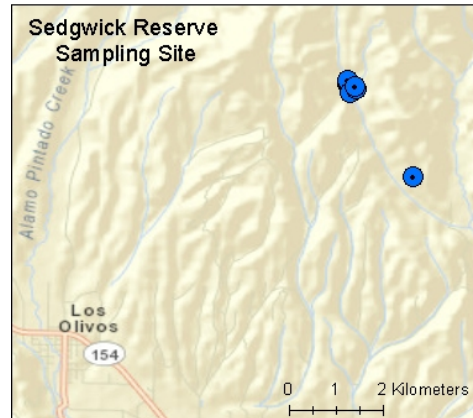
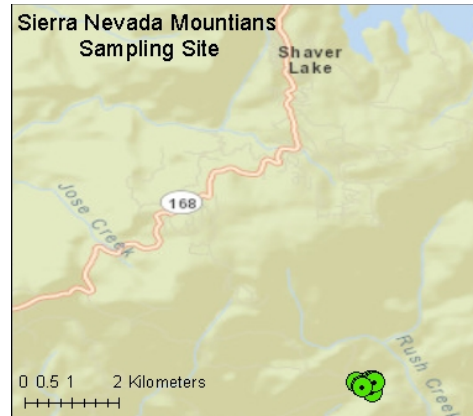
- Thermal Infrared (TIR) spectrum (2.5 to 15.4 μm) can answer some questions about plant characteristics
- Plants show spectral diversity in TIR
 - (Ullah et al., 2012; Ribeiro da Luz and Crowley 2010; Salisbury and Milton, 1988)
- Spectra features of cellulose, cutin, xylan, silica, and oleanolic acid present (Ribeiro da Luz and Crowley 2007)
- Increased resolution is increasing possibilities
- Integrating VSWIR and TIR
 - Limited knowledge on synergies
 - HyspIRI Preparatory Flights with AVIRIS and MASTER

Research Questions:

- Are biochemical properties of plant species distinguishable in the TIR spectrum?
- Do synergies between VSWIR and the TIR spectrum improve analysis of biochemical properties?

Study Sites:

- **Sierra Nevada Mountains:**
 - Elevation: 1400 m
- **Sedgwick Reserve:**
 - Elevation: 400 m
- **Santa Barbara Area:**
 - Elevation: 5 to 1080 m



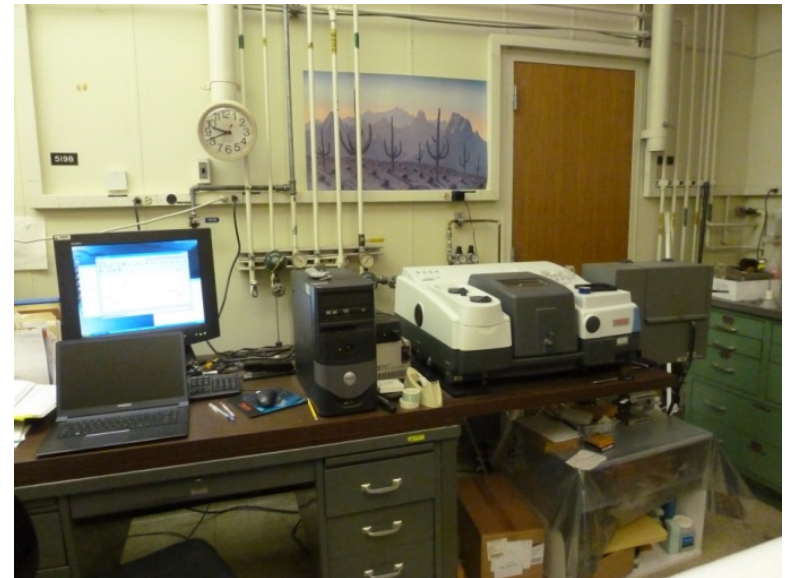
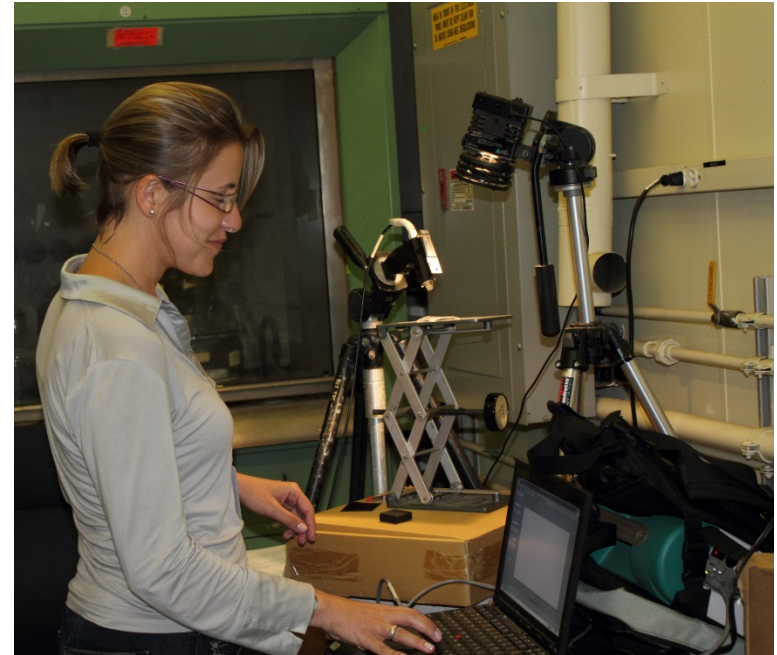
Sampling:

- 16 Species
- 3 Replications
- Young and Old Leaf Sample
- Spring:
 - Santa Barbara: 4/1/13
 - Sierra Nevada: 4/20/13
 - Sedgwick Reserve: 4/21/13
- Summer:
 - Santa Barbara: 6/3/13
 - Sierra Nevada: 6/8/13
 - Sedgwick Reserve: 6/9/13

<u>Species</u>	<u>Common Name</u>	<u>Abb.</u>
<i>Adenostoma fasciculatum</i>	Chamise	ADFA
<i>Arctostaphylos glandulosa</i>	Manzanita	ARGL
<i>Baccharis pilularis</i>	Coyote brush	BAPI
<i>Ceanothus cuneatus</i>	Buck-brush Ceanothus	CECU
<i>Ceanothus megacarpus</i>	Big-pod Ceanothus	CEME
<i>Ceanothus spinosus</i>	Green-bark Ceanothus	CESP
<i>Heteromeles arbutifolia</i>	Toyon	HEAR
<i>Umbellularia californica</i>	Bay laurel	UMCA
<i>Abies concolor</i>	White Fir	ABCO
<i>Pinus lambertiana</i>	Sugar Pine	PILA
<i>Pinus ponderosa</i>	Ponderosa Pine	PIPO
<i>Calocedrus decurrens</i>	Incense cedar	CADE
<i>Quercus agrifolia</i>	Coast live oak	QUAG
<i>Quercus douglasii</i>	Blue Oak	QUDO
<i>Quercus lobata</i>	Valley Oak	QULO
<i>Salvia leucophylla</i>	Purple Sage	SALE

Spectroscopy

- NASA Jet Propulsion Lab
- Thermal Spectrum
 - Nicolet
 - 2.5 μm to 15.4 μm
- VSWIR
 - Analytical Spectra Devices
 - 0.35 μm to 2.5 μm



Data Collected

- Lignin and Cellulose: Ankom Fiber Analyzer
- Nitrogen: Combustion Method using NA 1500 Nitrogen and Carbon analyzer
- Pigments: Processed by UC Davis*
- Specific Leaf Area*
- Leaf Thickness*
- Relative Water Content*

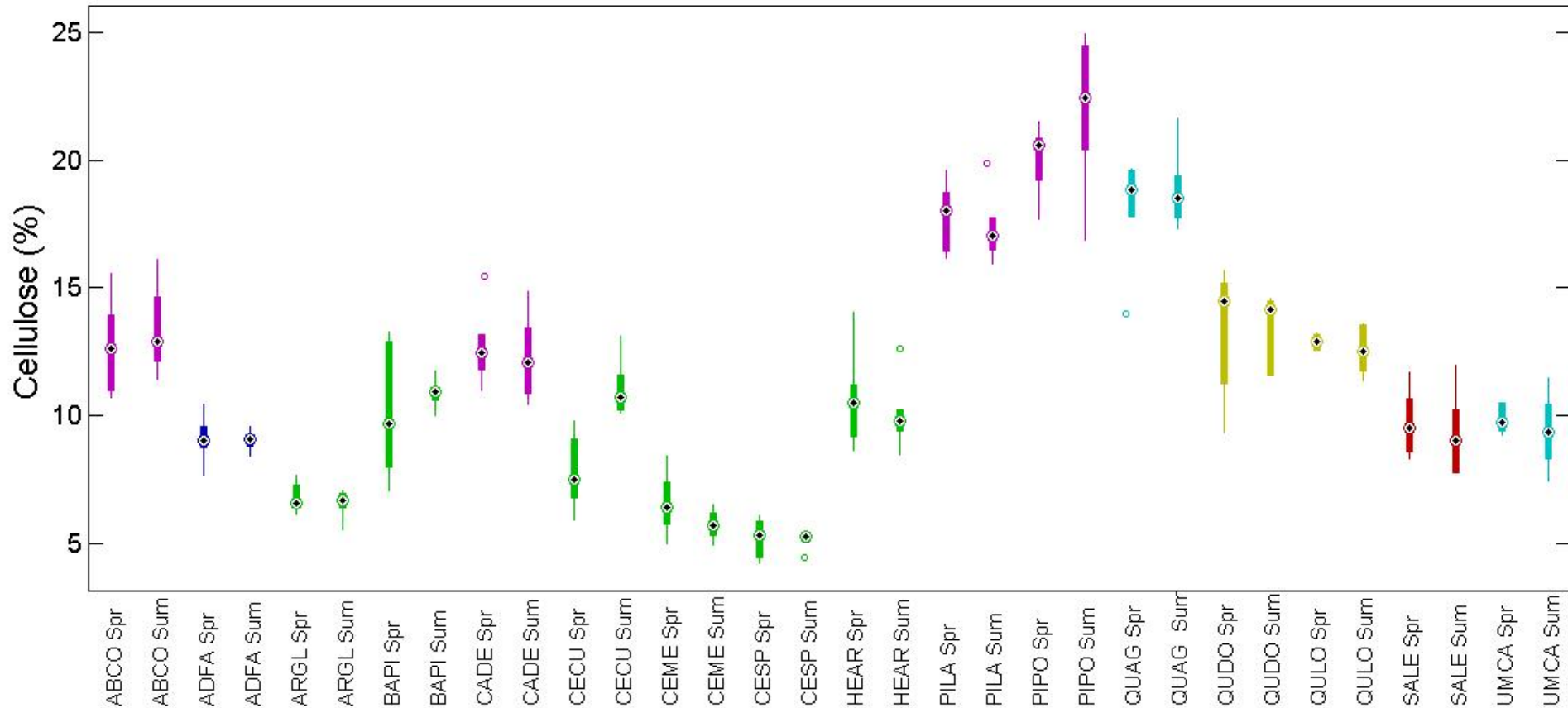
* Future Topics



PLS Regression

- Used in many spectroscopy studies
 - Nitrogen, Lignin, and Cellulose in the NIR (Bolster et al. 1996)
 - Leaf Mass Area, Nitrogen, Pigments, and Carotenoids in the VSWIR (Doughty et al., 2010)
 - Pigments, Carotenoids, LMA, Water, Nitrogen, Cellulose, Lignin, Tannins, Phosphorous, Potassium, and Zinc (Asner et al., 2011)
- Models a response variable when there are a large number of predictor variables
- Creates components as linear combinations of predictors
- Unique because response variables are taken into account
- Determine Number of Components:
 - Smallest RMSE
 - Highest Percent Variation Explained
 - Smallest Number of Components

Percent Cellulose By Season for All Species



Broadleaf Deciduous Shrub

Broadleaf Evergreen Tree

Broadleaf Deciduous Tree

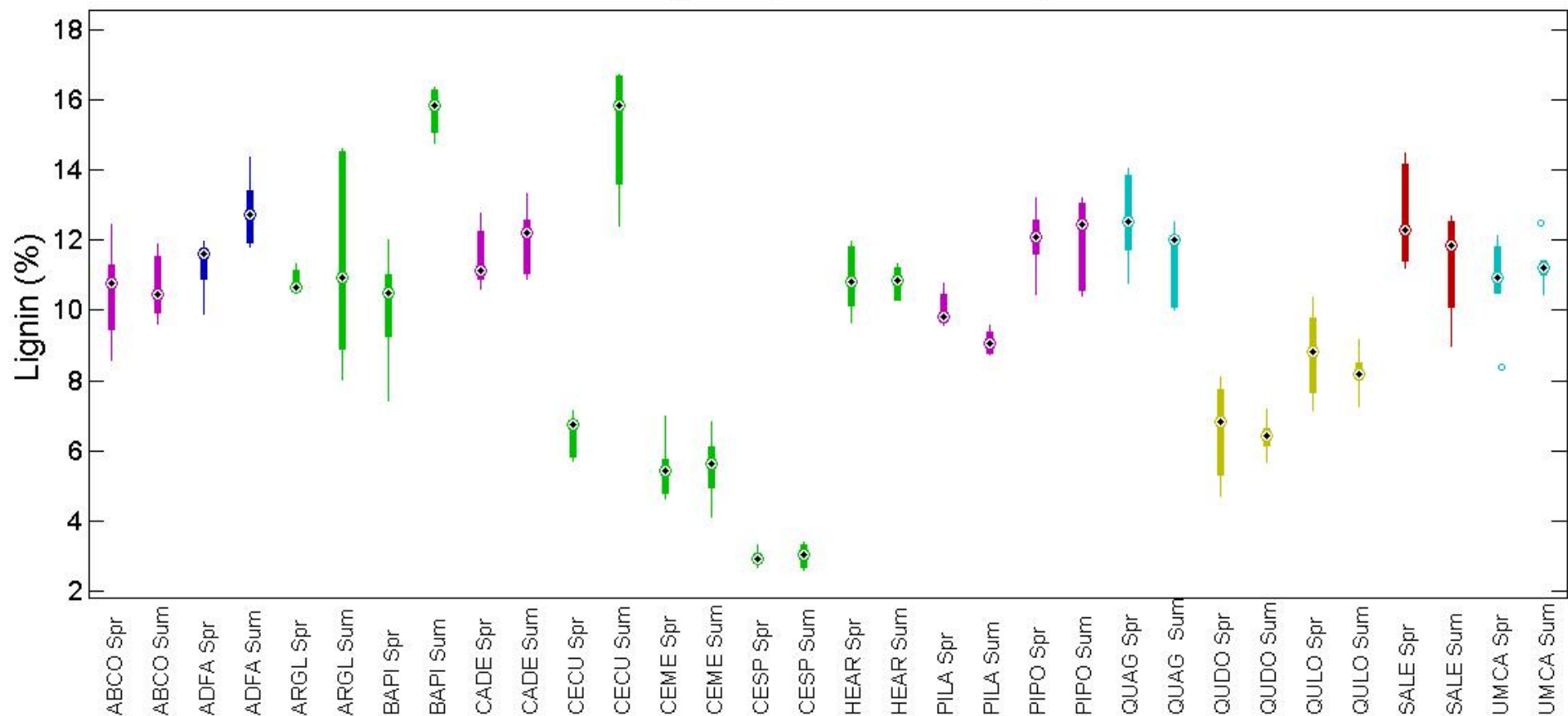
Needleleaf Evergreen Shrub

Broadleaf Evergreen Shrub

Needleleaf Evergreen Tree

Statistically Significant ($p < 0.05$) between Seasons: CECU

Percent Lignin By Season for All Species



Broadleaf Deciduous Shrub

Broadleaf Evergreen Tree

Broadleaf Deciduous Tree

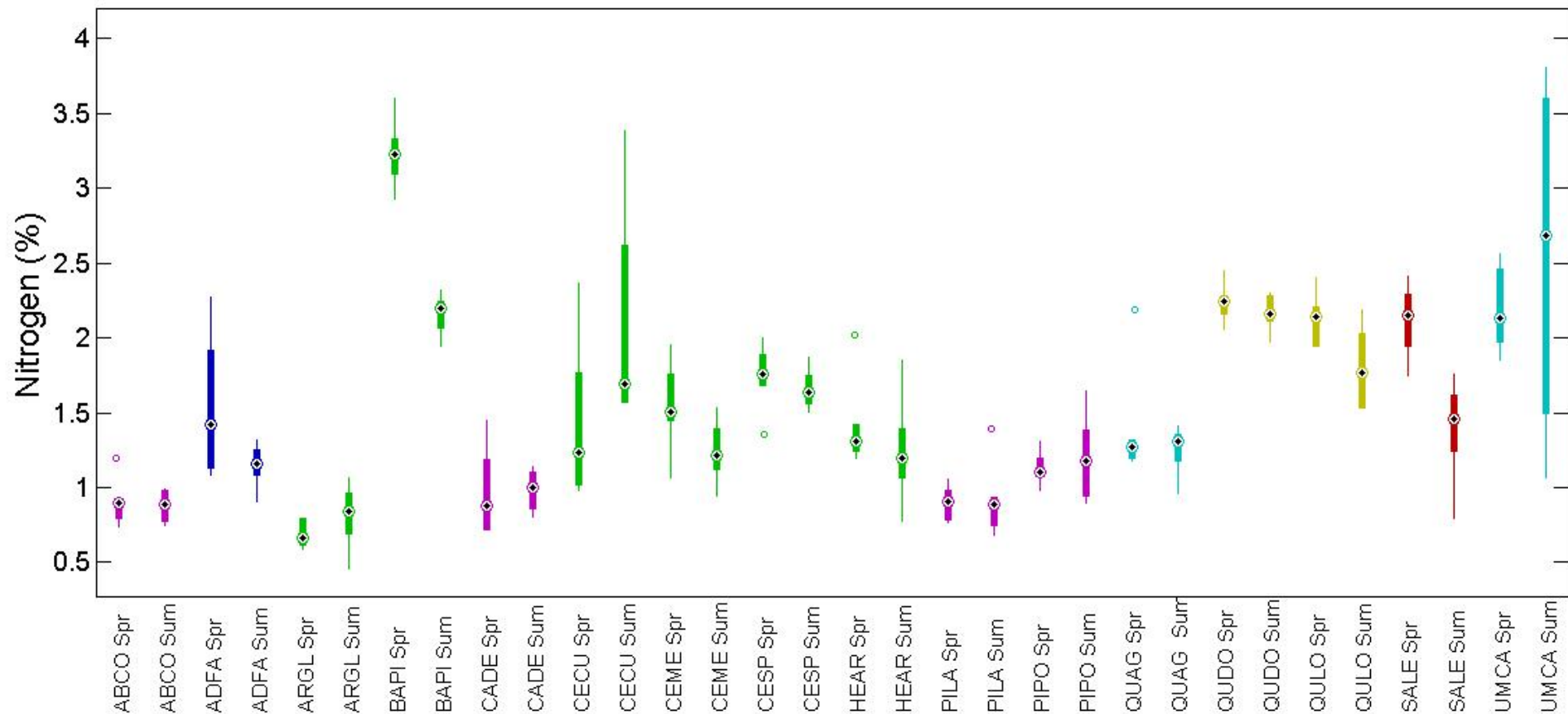
Needleleaf Evergreen Shrub

Broadleaf Evergreen Shrub

Needleleaf Evergreen Tree

Statistically Significant ($p < 0.05$) between Seasons: ADFA, BAPI, CECU, PILA

Percent Nitrogen By Season for All Species



Broadleaf Deciduous Shrub

Broadleaf Evergreen Tree

Broadleaf Deciduous Tree

Needleleaf Evergreen Shrub

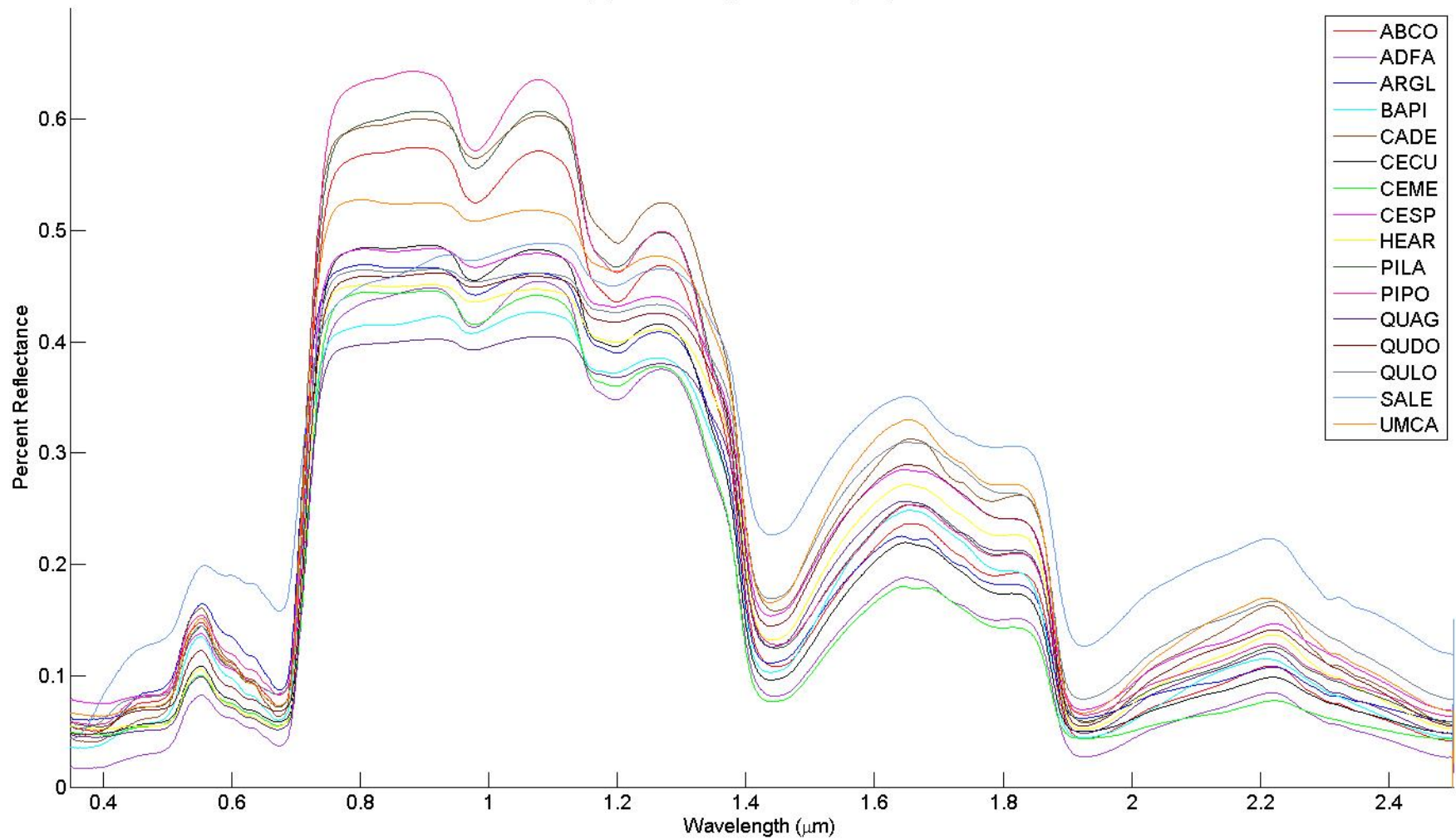
Broadleaf Evergreen Shrub

Needleleaf Evergreen Tree

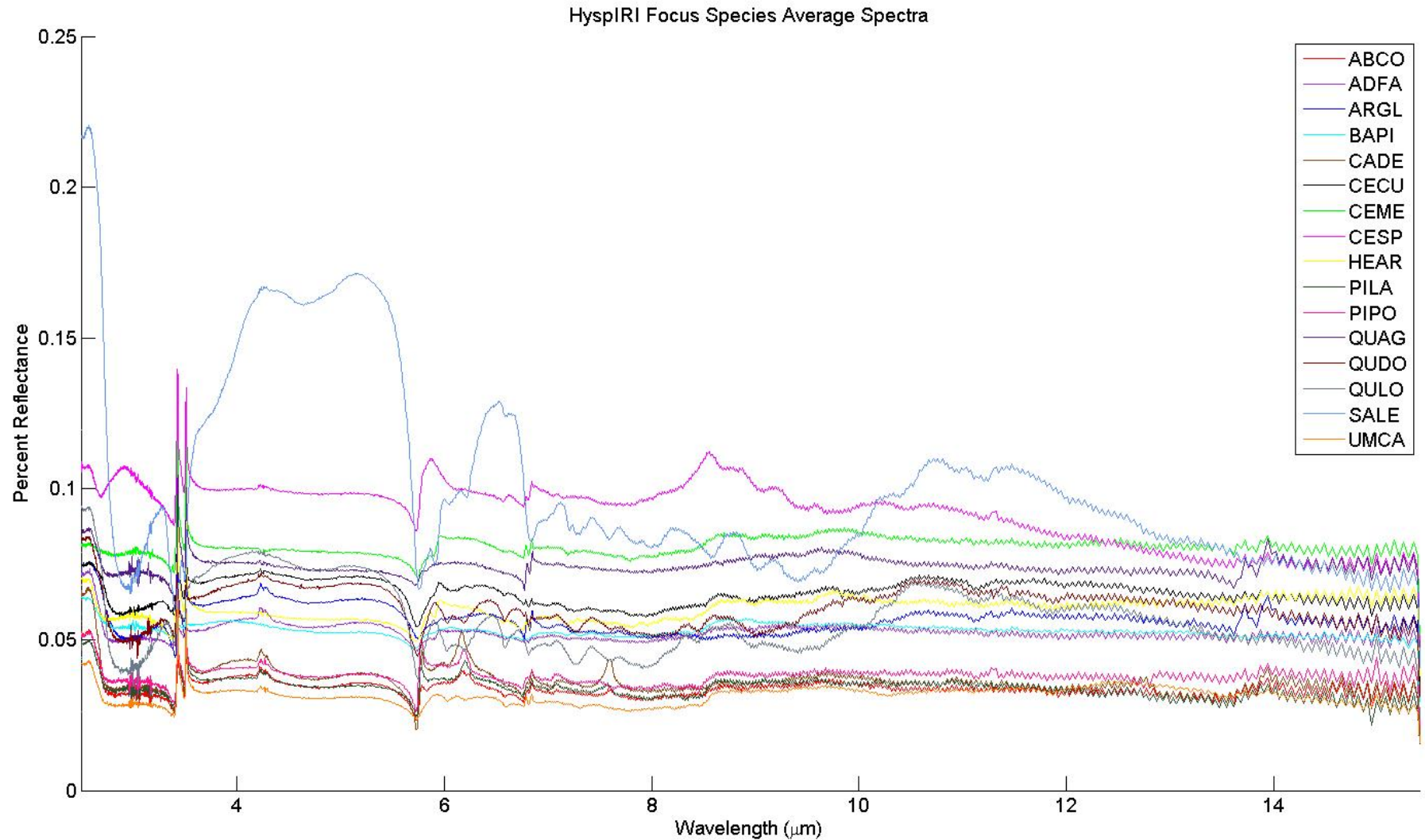
Statistically Significant ($p < 0.05$) between Seasons: BAPI, QULO, SALE

Spectra: VSWIR

HyspIRI Focus Species Average Spectra

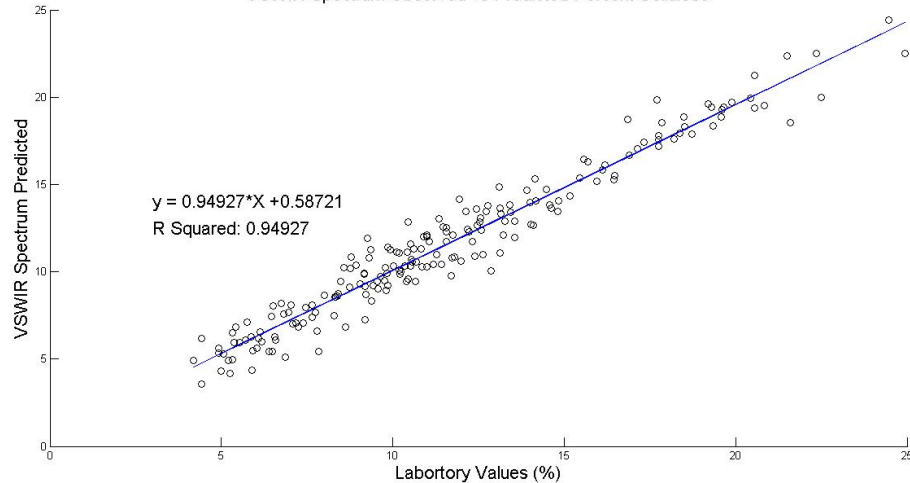


Spectra: TIR

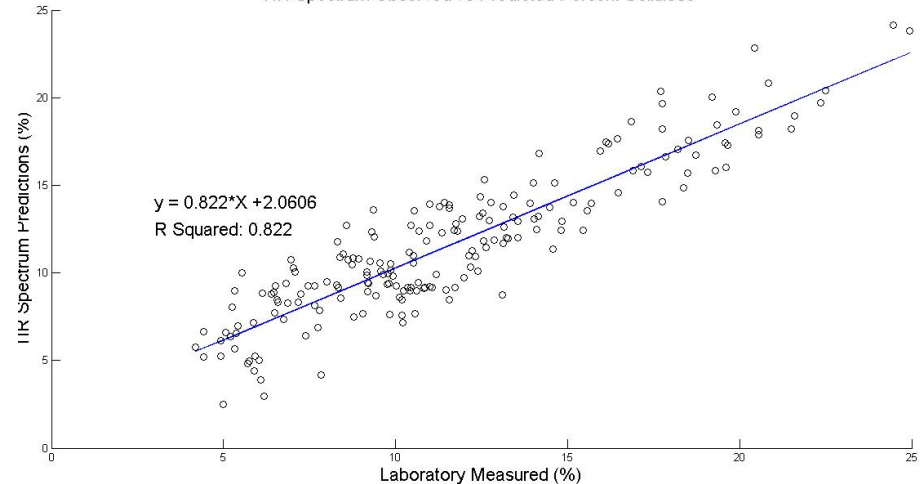


PLS Regression: Cellulose

VSWIR Spectrum Observed vs Predicted Percent Cellulose



TIR Spectrum Observed vs Predicted Percent Cellulose



Full Spectrum:

- 29 Components
- 99.1909 % Variance Explained

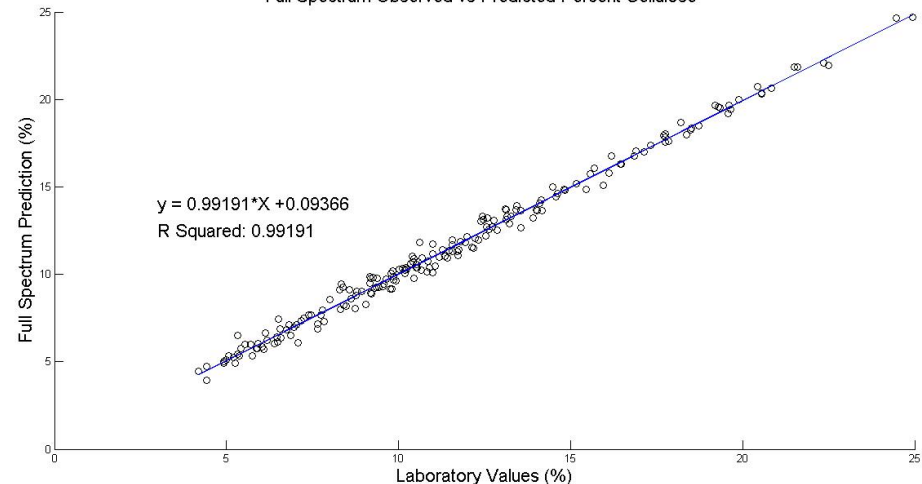
VSWIR:

- 27 Components
- 94.9274 % Variance Explained

TIR:

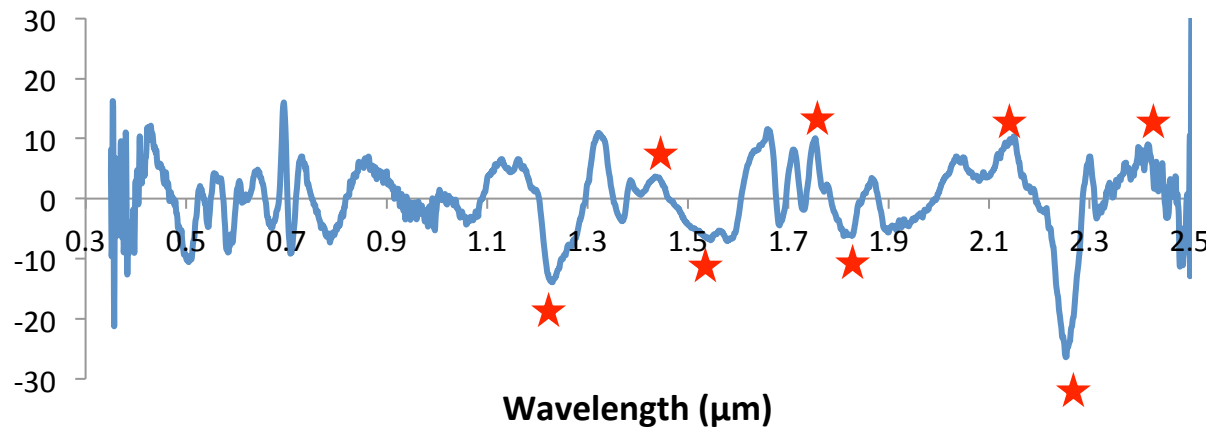
- 11 Components
- 82.1996 % Variance Explained

Full Spectrum Observed vs Predicted Percent Cellulose



PLS Regression: Cellulose

Cellulose Regression Coefficients for VSWIR

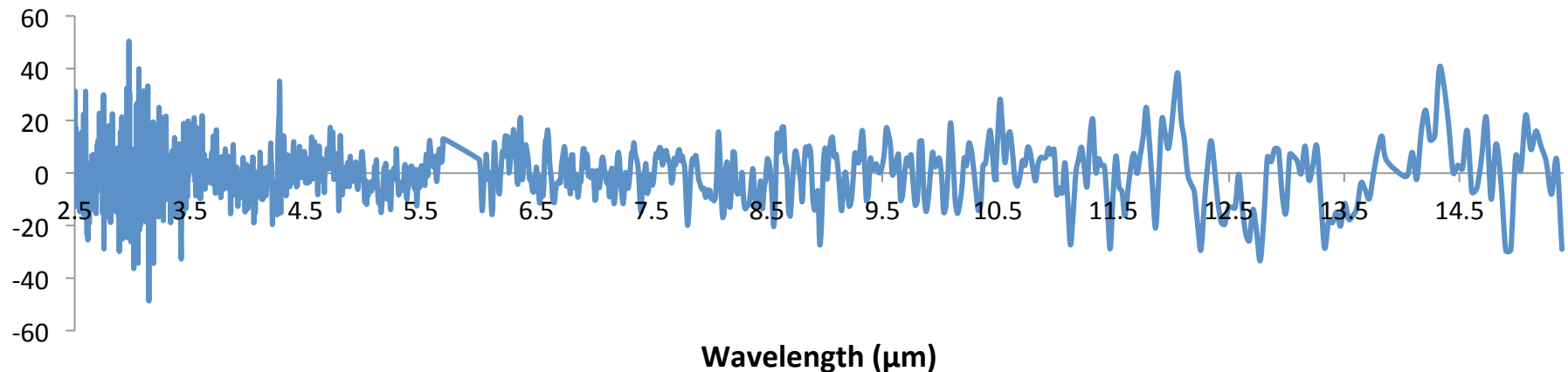


Wavelengths with Highest Coefficients

0.35	3.14
0.70	4.27
1.23	12.06
1.32	12.77
1.66	14.32
2.25	14.9
2.97	

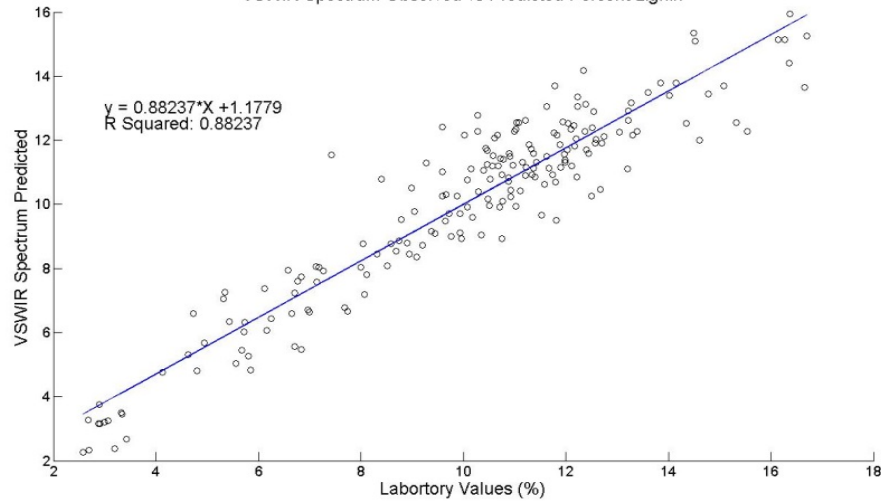
★ Known Absorption

Cellulose Regression Coefficients for TIR

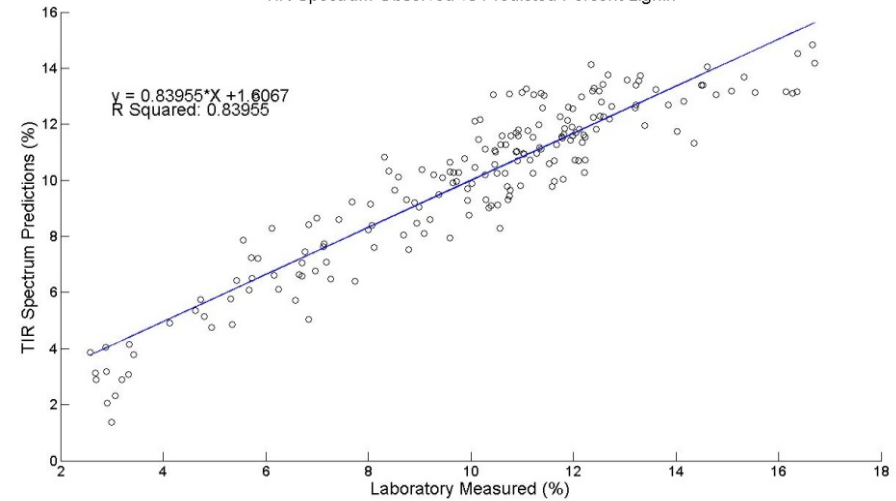


PLS Regression: Lignin

VSWIR Spectrum Observed vs Predicted Percent Lignin



TIR Spectrum Observed vs Predicted Percent Lignin



Full Spectrum:

- 27 Components
- 97.4780 % Variance Explained

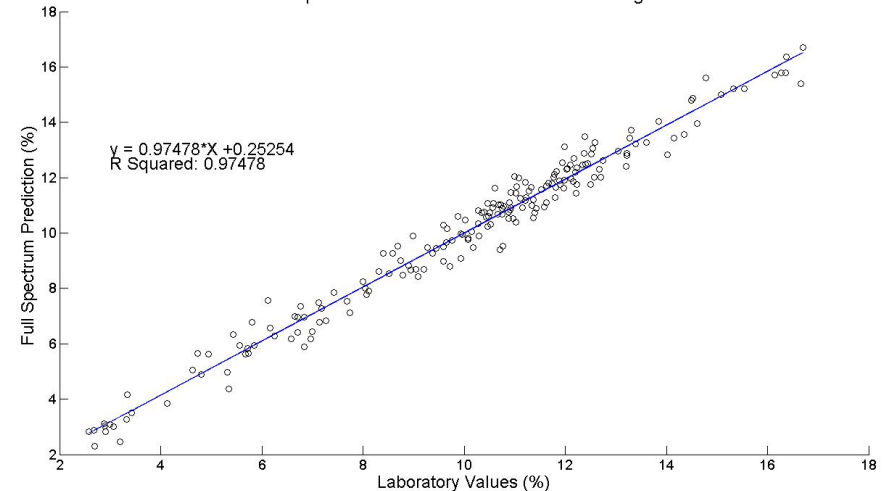
VSWIR:

- 25 Components
- 88.2368 % Variance Explained

TIR:

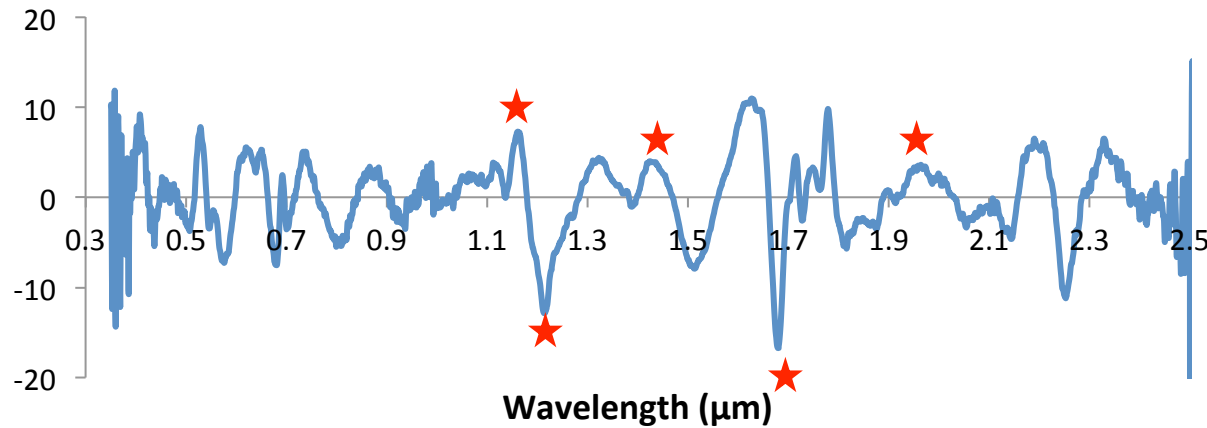
- 13 Components
- 83.9547 % Variance Explained

Full Spectrum Observed vs Predicted Percent Lignin



PLS Regression: Lignin

Lignin Regression Coefficients for VSWIR

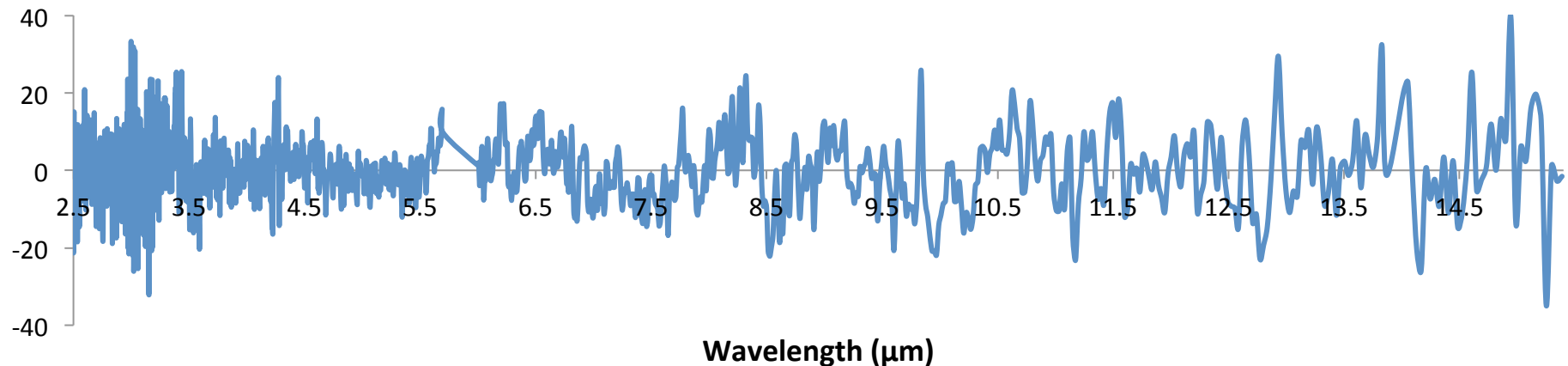


Wavelengths with Highest Coefficients

0.35	3.15
0.36	11.18
1.21	12.93
1.62	13.83
1.68	14.17
1.78	14.94
3.00	15.25

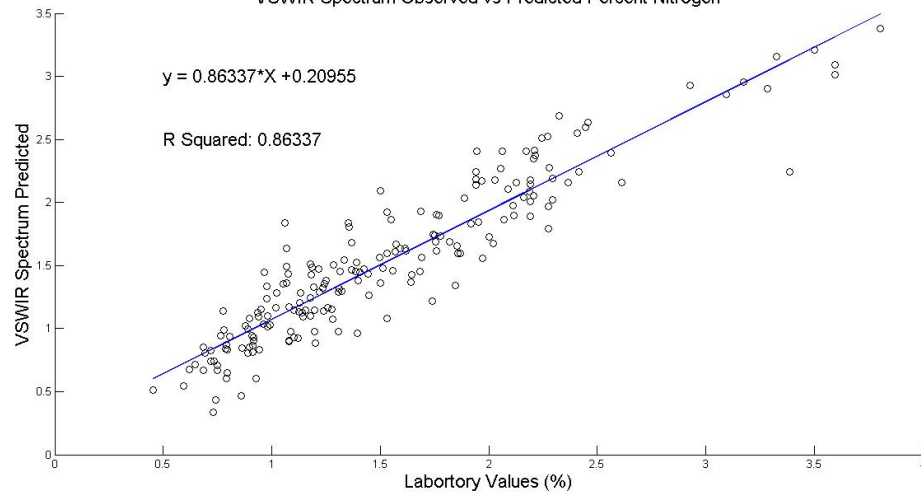
★ Known Absorption

Lignin Regression Coefficients for TIR

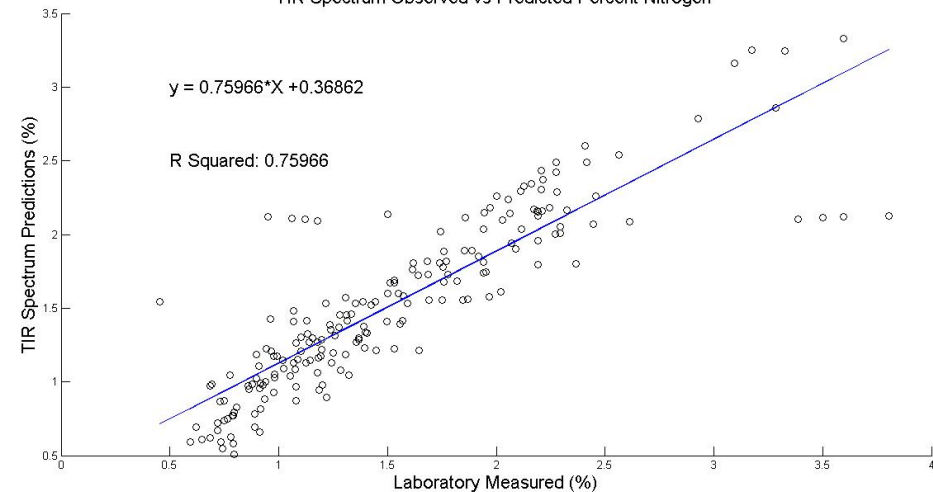


PLS Regression: Nitrogen

VSWIR Spectrum Observed vs Predicted Percent Nitrogen



TIR Spectrum Observed vs Predicted Percent Nitrogen



Full Spectrum:

- 17 Components
- 87.01 % Variance Explained

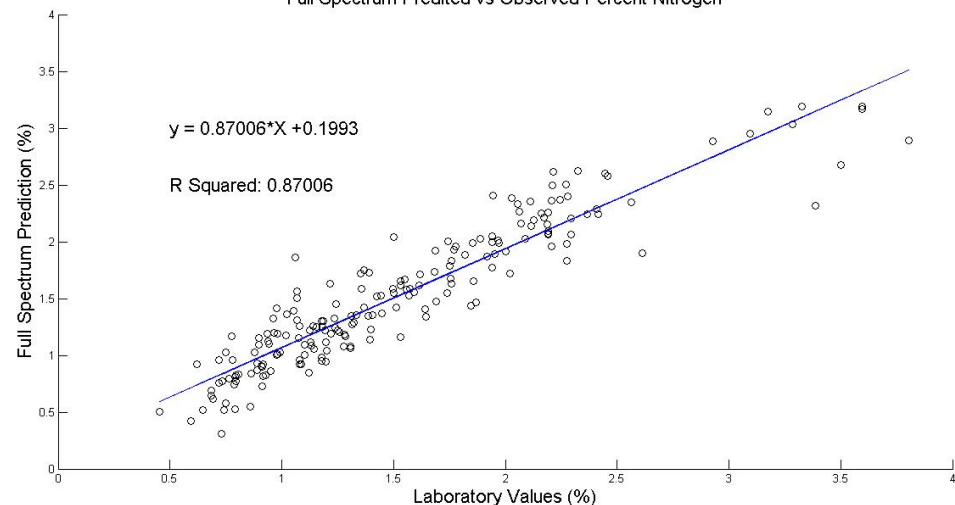
VSWIR:

- 19 Components
- 86.34 % Variance Explained

TIR:

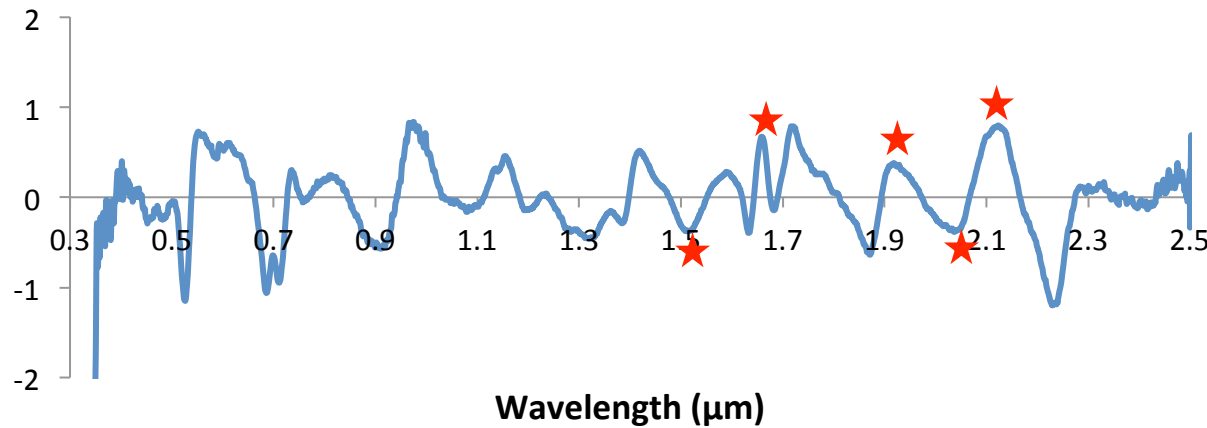
- 13 Components
- 75.97 % Variance Explained

Full Spectrum Predicted vs Observed Percent Nitrogen



PLS Regression: Nitrogen

Nitrogen Regression Coefficients for VSWIR

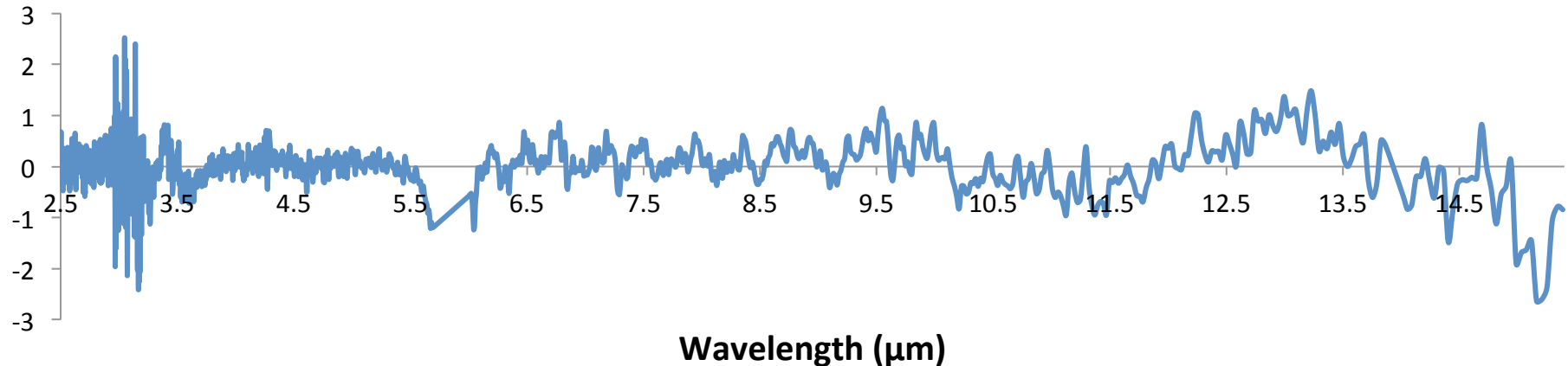


Wavelengths with Highest Coefficients

0.53	3.05
0.69	3.17
0.97	9.55
1.71	13.23
2.18	15.16
2.23	

★ Known Absorption

Nitrogen Regression Coefficients for TIR



Foliar Chemistry

- Seasonality changes in chemical components limited to only a few species
 - Different trend when Fall samples are collected?
 - Potentially use one regression equation for multiple seasons?
- Wide Range of Foliar Chemistry
 - Lignin: 2.6% to 16.7%
 - Cellulose: 4.2% to 24.9%
 - Nitrogen: 0.45% to 3.8%

PLS Regression

- Are biochemical properties of plant species distinguishable in the TIR spectrum?
 - Regressions were able to key into known absorption features of biochemicals in the VSWIR
 - TIR regression results had high R^2 , but not higher than VSWIR
- Do synergies between VSWIR and the TIR spectrum improve analysis of biochemical properties?
 - For all biochemicals the addition of TIR and VSWIR spectrum increased R^2

Summary

- Preliminary results from UCSB HyspIRI field campaign
- The incorporation of the TIR spectrum looks promising for foliar chemistry spectroscopy studies
- In the process of collecting Fall samples now
- In the future we will be looking at pigments, water content, specific leaf area, and leaf thickness

Questions?

References

- Asner, G.P., Martin, R.E., Ford, A.J., Metcalfe, D.J., and Liddell, M.J. 2009. Leaf chemical and spectral diversity in Australian tropical forests. *Ecological Applications* 19(1): 236-253.
- Asner, G.P. and Vitousek, P.M. 2005. Remote analysis of biological invasion and biogeochemical change. *Proceedings of the National Academy of Sciences of the United States of America* 102(12): 4383-4386.
- Asner, G. P., Wessman, C. A., Schimel, D. S., & Archer, S. (1998). Variability in leaf and litter optical properties: Implications for BRDF model inversions using AVHRR, Modis and MISR. *Remote Sensing of Environment* 63: 243–257.
- da Luz, B. R. and J. K. Crowley (2007). Spectral reflectance and emissivity features of broad leaf plants: Prospects for remote sensing in the thermal infrared (8.0-14.0 μm). *Remote Sensing of Environment* 109(4): 393-405.
- Bolster, K.L., Martin, L., Aber, M. 1996. Determination of carbon fraction and nitrogen concentration in tree foliage by near infrared reflectance: a comparison of statistical methods. *Canadian Journal of Forest Research* 26: 590-600.
- Ceccato, P., Flasse, S. Tarantola, S. Jacquemoud, S., Gregoire, J. 2001. detecting vegetation leaf water content using reflectance in the optical domain. *Remote Sensing of Environment* 77: 22-33.
- Daly C, Neilson RP, Phillips DL (1994) A statistical-topographic model for mapping climatological precipitation over mountainous terrain. *Journal of Applied Meteorology* 33: 140-158.
- Dennison, P.E., and Roberts, D.A. 2003. The effects of vegetation phenology on endmember selection and species mapping in southern California chaparral. *Remote Sensing Environment* 87(2-3): 123-135.
- Curran, P. J. (1989). Remote sensing of foliar chemistry. *Remote Sensing of Environment*, 30(3), 271–278. doi:10.1016/0034-4257(89)90069-2
- Doughty, C.E., Asner, G.P., Martin, R.E. 2009. Predicting tropical plant physiology from leaf and canopy spectroscopy. *Oecologia* 165(2): 289-299.
- Ferwerda, J. G., Skidmore, A. K., & Mutanga, O. (2005). Nitrogen detection with hyperspectral normalized ratio indices across multiple plant species. *International Journal of Remote Sensing*, 26 (18), 4083–4095. doi:10.1080/01431160500181044
- Garnier, E., Shipley, B., Rourmet, C., and Laurent, G. 2001. A standardized protocol for the determination of specific leaf area and leaf dry matter content. *Functional Ecology* 15(5): 688-695.
- Gitelson, A.A., Merzlyak, M.N., and Chivkunova, O.B. 2001. Optical properties and nondestructive estimation of anthocyanin content in plant leaves. *Photochemistry and Photobiology* 74(1): 38-45.
- Kokaly, R. F., Asner, G. P., Ollinger, S. V., Martin, M. E., & Wessman, C. a. (2009). Characterizing canopy biochemistry from imaging spectroscopy and its application to ecosystem studies. *Remote Sensing of Environment*, 113, S78–S91. doi:10.1016/j.rse.2008.10.018
- Kokaly, R.F. and Clark, R.N. 1992. Spectroscopic determination of leaf biochemistry using band-depth analysis of absorption features and stepwise multiple linear regression. *Remote Sensing of Environment* 67: 267–287.
- Martin, R.E., Asner, G.P., and Sack, L. 2007. Genetic variation in leaf pigment, optical, and photosynthetic function among diverse phenotypes of *Metrosideros polymorpha* grown in a common garden. *Oecologia* 151: 387- 400.
- Martin, M. E., Plourde, L. C., Ollinger, S. V., Smith, M.-L., & McNeil, B. E. (2008). A generalizable method for remote sensing of canopy nitrogen across a wide range of forest ecosystems. *Remote Sensing of Environment*, 112(9), 3511–3519. doi:10.1016/j.rse.2008.04.008
- Salisbury, J. W. and N. M. Milton (1988). Thermal infrared (2.5- μm to 13.5- μm) directional hemispherical reflectance of leaves. *Photogrammetric Engineering and Remote Sensing* 54(9): 1301-1304.
- Ullah, S., Schlerf, M., Skidmore, A.K., and Hecker, C. (2012). Identifying plant species using mid-wave infrared (2.5-6 μm) and thermal infrared (8-14 μm) emissivity spectra. *Remote Sensing of Environment* 118: 95-102.